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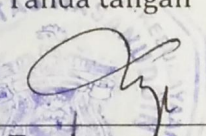
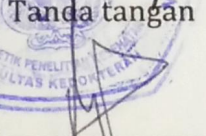
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REKOMENDASI PERSETUJUAN ETIK

Nomor : 681/UN4.6.4.5.31/ PP36/ 2019

Tanggal: 26 Agustus 2019

Dengan ini Menyatakan bahwa Protokol dan Dokumen yang Berhubungan Dengan Protokol berikut ini telah mendapatkan Persetujuan Etik :

No Protokol	UH19060344	No Sponsor Protokol	
Peneliti Utama	Dr.dr. Khairuddin Djawad, SpKK	Sponsor	
Judul Peneliti	Efek Protektif dan Terapeutik Formulasi Topikal (Krim, Gel, Serum) Ekstrak Kulit Manggis (Garcinia Mangostana L) terhadap Eritema Yang Diinduksi Oleh UVB		
No Versi Protokol	2	Tanggal Versi	15 Agustus 2019
No Versi PSP	2	Tanggal Versi	15 Agustus 2019
Tempat Penelitian	RSUP dr. Wahidin Sudirohusodo dan RS Universitas Hasanuddin Makassar		
Jenis Review	<input type="checkbox"/> Exempted <input type="checkbox"/> Expedited <input checked="" type="checkbox"/> Fullboard Tanggal 3 Juli 2019	Masa Berlaku 26 Agustus 2019 sampai 26 Agustus 2020	Frekuensi review lanjutan
Ketua Komisi Etik Penelitian Kesehatan FKUH	Nama Prof.Dr.dr. Suryani As'ad, M.Sc.,Sp.GK (K)	Tanda tangan 	
Sekretaris Komisi Etik Penelitian Kesehatan FKUH	Nama dr. Agussalim Bukhari, M.Med.,Ph.D.,Sp.GK (K)	Tanda tangan 	

Kewajiban Peneliti Utama:

- Menyerahkan Amandemen Protokol untuk persetujuan sebelum di implementasikan
- Menyerahkan Laporan SAE ke Komisi Etik dalam 24 Jam dan dilengkapi dalam 7 hari dan Laporan SUSAR dalam 72 Jam setelah Peneliti Utama menerima laporan
- Menyerahkan Laporan Kemajuan (progress report) setiap 6 bulan untuk penelitian resiko tinggi dan setiap setahun untuk penelitian resiko rendah
- Menyerahkan laporan akhir setelah Penelitian berakhir
- Melaporkan penyimpangan dari protokol yang disetujui (protocol deviation / violation)
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Protective Effect of Mangosteen Pericarp Extract Cream Against UVB-Induced Erythema

Article in *Journal of Clinical and Aesthetic Dermatology* · October 2020

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ABSTRACT

BACKGROUND: Exposing the skin to ultraviolet B (UVB) radiation triggers inflammation, with erythema as the most prominent acute clinical manifestation. Xanthones, secondary metabolites of the mangosteen pericarp, have been shown to possess anti-inflammatory and antioxidant properties. With the increasing evidence of harmful effects of chemical sunscreens to marine organisms, it is necessary to develop environmentally friendly sunscreen options.

OBJECTIVE: We sought to assess the protective effect of a nano-liposomal mangosteen pericarp extract (MPE) cream against UVB-induced erythema. **METHODS:** Thirty-one healthy subjects with Fitzpatrick Skin Types III or IV were enrolled. Six sites on the back of each volunteer were used for testing; Sites 1 to 4 were exposed to UVB at a dosage of two minimal erythema dose. Before UVB exposure, the test materials, including 5%, 10%, or 20% MPE cream, or a base cream without MPE, were applied to each site. Sites 5 and 6 served as the positive (UVB only, no treatment) and negative (no treatment, no UVB) control groups. The lightness index (ΔL^*) and erythema index (Δa^*) were assessed before and 24 hours after treatment using Chromameter. **RESULTS:** On all treated sites, there were only slight decrease of ΔL^* and increase of Δa^* . The ΔL^* of the 5%, 10%, and 20% cream groups were 1.65, -0.4, and 0.01, respectively, which were significantly different from UVB group (4.53). The Δa^* of the 5%, 10%, and 20% cream (1.03, -0.07; 0.16, respectively) was significantly different from the base cream (2.03) and UVB group (5.585). **CONCLUSION:** The 10% and 20% nano-liposomal cream of MPE appeared to exert protective effects against UVB-induced erythema on human skin.

KEY WORDS: Chromameter, erythema, mangosteen pericarp extract, sunscreen, ultraviolet B

Protective Effect of Mangosteen Pericarp Extract Cream Against UVB-Induced Erythema

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The skin occupies a strategic location between the harmful external environment and the biochemically active internal environment. Ultraviolet (UV) radiation is an external environmental factor that can harm the skin. Excessive sun exposure can cause erythema, edema, sunburn, hyperplasia, premature aging, melanoma, and non-melanoma skin malignancies.¹ Erythema is the most visually acute prominent aspect of the sunburn response. The inflammatory process manifests as erythema and edema due to increased vasodilation and vascular permeability.² Erythema appears in a diphasic pattern, usually seen 3 to 5 hours after exposure, then reaches maximum intensity after 12 to 24 hours and generally subsides within 72 hours. Various biochemical processes at the cellular level occur, including increased blood flow, endothelial cell activation, and increased levels of inflammatory mediators.³ The absorbed UVB radiation activates the enzyme phospholipase and releases arachidonic acid from cell membrane phospholipids. Arachidonic acid is then converted to prostaglandin (PGE) and 12-hydroxyeicosatetraenoic acid (12-HETE) by the enzymes cyclooxygenase (COX) and lipoxygenase (LOX).² UVB exposure triggers inflammation through direct induction of epidermal keratinocytes to produce proinflammatory cytokines, including TNF- α ,

IL-1 β , and COX-2.⁴

Mangosteen (*Garcinia mangostana* L.), is mainly cultivated in Indonesia, Malaysia, the Philippines, and Thailand.⁵ Secondary metabolites of mangosteen pericarp, called xanthones, have broad pharmacological properties, including anti-inflammatory, antineoplastic, antioxidant, antiproliferation, anticancer, antimalarial, antibacterial, antiobesity, hepatoprotective, neuroprotective, and cardioprotective.^{6,7} A total of 68 organic xanthone compounds have been isolated from all parts of the mangosteen.⁸ The highest xanthones concentration, reaching 78, has been isolated from the mangosteen pericarp, namely α -mangostin.^{9,10} Studies have shown α -mangostin can inhibit the production of inflammatory mediators and decrease levels of mRNA expression from TNF- α , IL-1 β , and IL-6.⁴ Also, MPE has been shown to inhibit COX enzymes in the arachidonic acid pathway.¹¹

Although current evidence suggests that the use of sunscreens does not yield any adverse health effects, the potentially harmful environmental effects of sunscreen agents are still of concern. Currently, there are issued statements in certain areas of the United States prohibiting sunscreens containing oxybenzone and octinoxate, since these materials have been shown to have a fatal impact on the maintenance

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CORRESPONDENCE: Khairuddin Djawad, MD, PhD; Email: duddin@gmail.com

TABLE 1. Ingredients, suppliers, and composition (%weight) of the cream base, 5%, 10%, and 20% MPE cream used in this study

COMPONENTS (INCI)	SUPPLIERS	5% CREAM	10% CREAM	20% CREAM	CREAM BASE
		(%W/W)			
GML pericarp extract / MPE	Faculty of Pharmacy Hasanuddin University	5	10	20	-
Ethanol 96%	Dianum (Emsurf)	5	5	5	5
PEG-40 HCO	Brataco Chemical (KAO)	5	5	5	5
Cetyl alcohol	Multikimia Raya (Croda Oleo)	5	5	5	5
Paraffin liquidum	Brataco Chemical	3	3	3	3
Isopropyl Myristate	Jerindo Pratama (Croda Oleo)	5	5	5	5
Sorbitan stearate	Multikimia Raya (Croda Oleo)	3,5	3,5	3,5	3,5
Stearic acid	Brataco Chemical (Croda Oleo)	3,5	3,5	3,5	3,5
Cetearyl alcohol	Multikimia Raya (Croda Oleo)	2	2	2	2
TEA	Jerindo Pratama	0,2	0,2	0,2	0,2
Dimethicone	Tristar Chemical (KAO)	1	1	1	1
Sorbitol	Brataco Chemical (KAO)	5	5	5	5
Methyl paraben	Brataco Chemical	0,2	0,2	0,2	0,2
Sorbic acid	Multikimia Raya	0,5	0,5	0,5	0,5
Titanium dioxide	Multikimia Raya	0,5	0,5	0,5	0,5
Aqua demineralisata	UBAYA	46,6	41,6	31,6	60,6

INCI: International Nomenclature of Cosmetic Ingredients; GML: *Garcinia mangostana*; HCO: Hydrogenated castor oil

of coral reefs, which play an important role in supporting the preservation of fish resources and marine organisms.¹² Therefore, it is necessary to develop safer natural ingredients as sunscreen agents, such as from plant extracts that contain photoprotective and antioxidant effects.¹³

This double-blind, randomized, controlled study was designed to clinically assess the protective effect of a nano-liposomal MPE cream against UVB-induced erythema on human skin.

METHODS

Subjects. Thirty-one healthy subjects with Fitzpatrick Skin Types III or IV were enrolled in the study. All participants followed the study's specific protocol and signed an informed consent form. The subjects were 20 to 45 years of age with no history of photodermatitis or skin cancer and were not taking any photosensitizing or anti-inflammatory medications. This study complied with the ethical guidelines and was approved by the Health Research Ethics Commission Committee of Hasanuddin University (process No. 681/UN4.6.4.5.31/PP36/2019).

Materials. Mangosteen pericarp liquid extract was made at the laboratory of Phytochemistry and Pharmacognosy, Faculty of Pharmacy at Hasanuddin University. The MPE creams were prepared at the Laboratory of Pharmaceutical Technology, Faculty of Pharmacy

at the University of Surabaya (UBAYA), based on the ingredients listed in Table 1. Four formulas were created using the same cream base; the first formula comprised the cream base without active ingredients, the second formula contained 5% mangosteen pericarp liquid extract, the third formula contained 10% mangosteen pericarp liquid extract, and the fourth formula contained 20% mangosteen pericarp liquid extract.

Preparation of skin cream. The cream was formulated by melting the oil phase (cetyl alcohol, sorbitan stearate, Cetearyl alcohol, isopropyl myristate, paraffin liquidum, stearic acid) at 65 °C. The water phase (aqua, sorbitol, methylparaben, sorbic acid, and TEA) was heated at 65 °C. Then, the water phase was put into the oil phase and stirred with Ultraturrax at a speed of 11.000rpm for three minutes, then the stirring speed was reduced to 3.000rpm and stirred for another five minutes. Stirring was continued for five minutes, then dimethicone and titanium dioxide were added, then stirring continued for five minutes to form a homogeneous emulsion. MPE was dissolved in 96 ethanol in a mortar, then added PEG-40 HCO, crushed and stirred until a homogeneous mixture was formed. The extract mixture was then put into the emulsion while still stirring at 3.000rpm for three minutes, then cooled while still stirring.

Accelerated stability study. To obtain

stability data of the MPE cream at all formulas, an accelerated stability study was conducted for six months using the climatic chamber with a temperature of 25 to 28 °C and relative humidity of 60 to 70 percent. Parameters that were measured include organoleptics, pH, and viscosity. Data were recorded every month.

Solar simulator. Psora-Comb Dermalight 80 (Dr. K. Honle GmbH, Munich, Germany) was used as a UVB emitting source. It emits a continuous spectrum of UVB radiation, with peaks of 313nm, placed at a fixed distance of 3cm using its comb attachment.

Determination of the Minimal Erythema Dose (MED). Once the skin type was defined using Chromameter CR-400 (Konica Minolta, Inc., Tokyo, Japan) and Individual Typology Angle (ITA), the determination of the MED was performed at the first visit. Daavlin MED dose patch (Daavlin Company; Bryan, Ohio) was used on the back of each, which comprises six 1.9x1.9 cm squares (3.61 cm² area). The MED is defined as the lowest dose that induces erythema in an individual.¹⁴ The MED is determined by exposing six squares to gradually increasing amounts of UVB radiation (200, 400, 600, 800, 1000, 1200mJ/cm²) and examining the UV-exposed areas 24 hours later.¹⁵ The area with the least erythema observed by the investigators is considered the MED subjectively, while

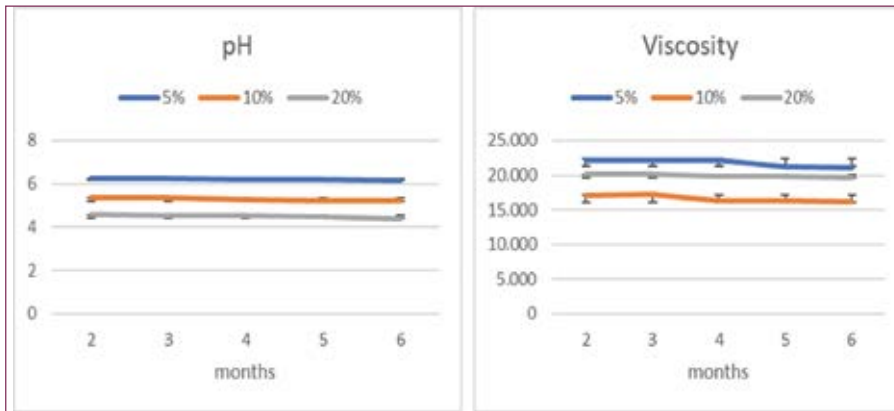


FIGURE 1. The pH and viscosity of the 5%, 10%, and 20% MPE creams remained stable for six months in storage in a climatic chamber kept at 25–28°C with a relative humidity of 60–70%

objectively we used a 2.5-point difference in the a^* value before and after irradiation to indicate a significant difference in erythema.¹⁶ Thus, in this study, the determination of the MED was performed subjectively and objectively.

Treatment protocol. This study was conducted in our dermatology and venereology outpatient clinic in September 2019. Before each irradiation, the following materials were applied to sites 1 to 4, respectively: 5% MPE cream, 10% MPE cream, 20% MPE cream, and cream base. No product was applied to Site 5, which served as the positive control, and neither UVB nor MPE cream was applied to Site 6, which served as the negative control. A standardized dose of 2.77mg/cm² MPE cream was calculated, in which 10mg of MPE cream was applied to each site. All test product applications were performed by other volunteers using latex gloves to maintain the double-blind study. The MPE cream was applied and left for 20 minutes before irradiation. A summary of the treatment sites is given in Table 2.

We chose to evaluate the creams on the back of each participant, as it proved to be the test site with the smallest fluctuations of photometric measurements.¹⁷ Six squares of the Daavlin MED dose patch was used on the back of each subject with designation as Sites 1 to 6. All subjects received the equivalent of 2 MED in each site. This dose is considered sufficient to induce defined erythema that can be separated from the background readings of untreated.^{17,18} Irradiation was performed at the second visit, then chromameter measurements were performed after 24 hours to measure the erythema and lightness index.

Chromameter measurement. Before

and 24 hours after exposure to UVB, the L^* and a^* values on the test sites were measured using a Chromameter CR-400 (Konica Minolta, Inc., Tokyo, Japan). The values were measured three times on each test site, and the mean values were determined. The differences (Δ) in median L^* and a^* values before and 24 hours after irradiation were calculated. Based on the Commission Internationale de l'Éclairage (CIE) system, the output values are expressed in three dimensions (L^* , a^* , b^*), where a^* represents erythema and L^* is to assess lightness. The erythema index (Δa^*) and lightness index (ΔL^*) was assessed by a difference (Δ) in baseline and 24 hours after treatment.

Statistical analysis. The data were analyzed using SPSS 25.0 software (SPSS Inc., Chicago, Illinois). Nonparametric statistical methods (Kruskal Wallis and Mann-Whitney test) were used with P -value <0.05 was considered statistically significant.

RESULTS

The cream stability. A difference in the pH of the three creams is caused by the differences in extract concentration. The pH of the cream decreases as the concentration of the MPE increases. Over a period of six months in storage, no significant pH differences were observed in each of the creams. Likewise, the cream viscosity did not show a significant difference over six months of storage (Figure 1). Changes in viscosity are also influenced by differences in extract concentration. The lowest viscosity was observed in the 10 percent MPE cream and the highest was observed in the five percent MPE cream. The pH and viscosity in the 5 percent, 10 percent, and 20 percent MPE creams were found to be stable

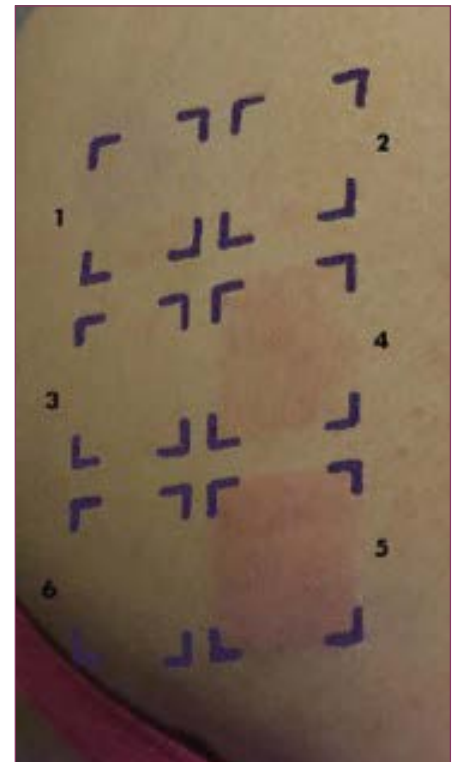


FIGURE 2. Clinical photograph of a study participant; Site 1: 5% mangosteen pericarp extract (MPE) cream+UVB; Site 2: 10% MPE cream+UVB; Site 3: 20% MPE cream+UVB; Site 4: Base Cream+UVB; Site 5: UVB only (positive control); Site 6: No treatment, no UVB (negative control). On Sites 1–3, minimal to no erythema was observed.

TABLE 2. Summary of the treatment groups in this study; the mangosteen pericarp extract (MPE) cream and the base cream was applied for 20 minutes before 2 minimal erythema dose UVB irradiation on the subject's trunk (each square of 3.61 cm² area)

SITE	TREATMENT
1	5% MPE cream, then UVB
2	10% MPE cream, then UVB
3	20% MPE cream, then UVB
4	Base cream, then UVB
5	UVB only (positive control)
6	No treatment, no UVB (negative control)

while in storage for six months in a climatic chamber at 25 to 28 °C and a relative humidity of 60 to 70 percent.

Clinical photographs. After UVB irradiation, some irradiated sites developed some degrees of erythema, as shown in Figure 2. Obvious erythema was seen on Site 5 (UVB) and Site 4 (cream base + UVB), suggesting the cream base alone provided minimal protection from UVB.

The erythema index. All subjects showed significant differences in the erythema index

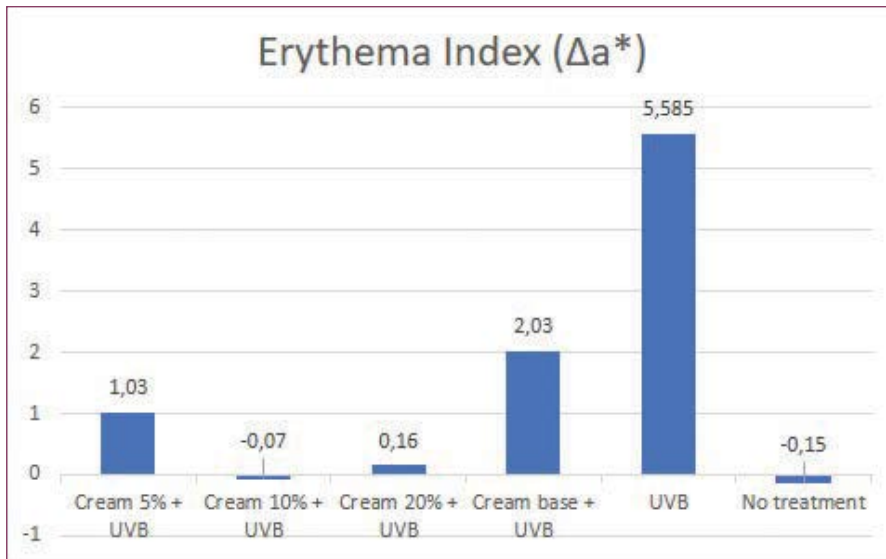


FIGURE 3. The Erythema Index (Δa^*) of each treatment group ($a^*_{24\text{ hours}} - a^*_{\text{baseline}}$)

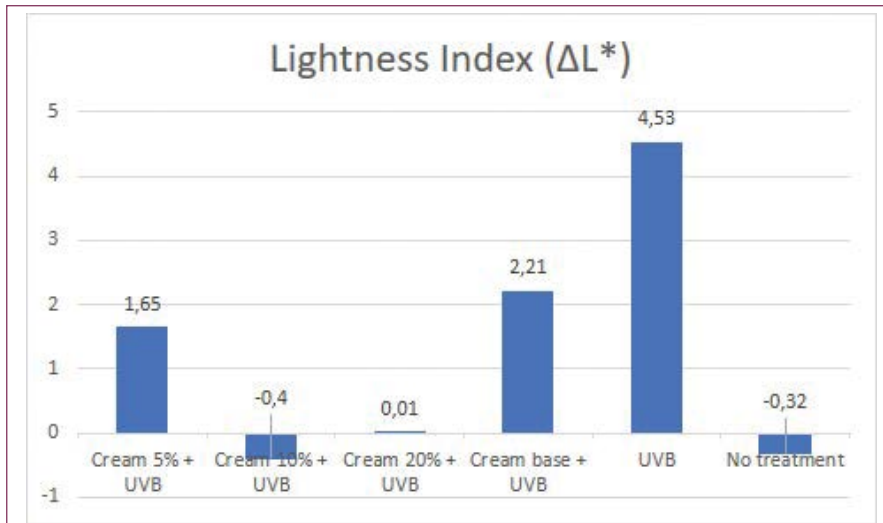


FIGURE 4. The Lightness Index (ΔL^*) of each treatment group ($L^*_{24\text{ hours}} - L^*_{\text{baseline}}$)

in the UVB and no treatment group (Figure 3). This is consistent with a statistically significant P -value ($p < 0.05$) between both treatment groups with Δa^* values of 5.585 and -0.15, respectively (Table 3). Based on this observation, 2 MED exposure to UVB was shown to cause significant erythema. Overall, the administration of MPE cream was found to reduce the erythema index. This is consistent with the statistically significant difference in the erythema index among the six treatment groups ($p < 0.05$). The erythema index (Δa^*) of the 5%, 10%, 20% cream (1,03; -0,07; 0,16) differed significantly ($p < 0.05$) to base cream (2,03) and to UVB group (5,585). However, the erythema index of the base cream was also significantly different from

the UVB group ($p < 0.05$), indicating a minimal protective effect against UVB. Based on the concentration ratio, the lowest erythema index was found in the 10% cream group followed by the 20% cream group and the 5% cream group. Statistical data supports the superiority of the 10% and 20% creams over the 5% cream, whereas the 10% and 20% creams did not differ significantly ($p = 0.198$). The 10% cream group and the 20% cream group were not significantly different when compared to the no treatment group, with values of Δa^* -0.07 ($p = 1.000$) and 0.16 ($p = 0.210$), respectively. This finding indicates that the 10% cream and 20% cream exhibit a substantial protective effect against UVB-induced erythema equivalent to the no

treatment group.

The lightness index. The lightness index in the UVB and no treatment group showed a significant difference ($p < 0.05$) as shown in Table 4, with each L^* value of 4.53 and -0.32, respectively, demonstrating 2 MED UVB exposure can significantly influence the lightness of the skin's color (Figure 4). Overall, the administration of MPE cream was found to reduce the lightness index (ΔL^*). This is consistent with the statistically significant difference between the six treatment groups ($p < 0.05$). The lightness index of the 5%, 10%, and 20% cream groups differ significantly to UVB group ($p < 0.05$), with each value ΔL^* 1.65; -0.4; 0.01, respectively. Moreover, the 10% and 20% cream groups were significantly different from base creams ($p < 0.05$), but not 5% cream ($p = 0.314$). These findings indicate that the base cream also exerts some protective effect against UVB-induced erythema. Although the 10% and 20% cream groups showed better protection, the base cream and the 5% cream group showed a similar protective effect. Based on three concentrations, the 10% cream was found to be more superior, followed by the 20% cream and the 5% cream group, though the 10% cream and the 20% cream were not significantly different ($p = 0.451$). When compared to the no treatment group, the 10% and 20% creams did not differ significantly, with a P -value of 0.746 and 0.410, respectively. These findings exhibit the protective ability of MPE cream that was found to be quite similar to the no treatment group based on the lightness index evaluation.

DISCUSSION

The Fitzpatrick Skin Type I to VI is the most useful clinical scale in assessing skin sensitivity to UV radiation, with lesser skin types more susceptible to sunburn and at a higher risk of developing skin cancer.¹⁹ This study was conducted on 31 subjects with Fitzpatrick Skin Types III and IV, which constitute the majority of the skin color of the Indonesian population. The sensitivity of an individual's skin to UV radiation can be measured by visual assessment of a MED. In general, MED increases with skin type.²⁰ Most studies have used twice the MED to induce defined erythema.^{17,18}

The peak intensity of UVB radiation occurring during the day can have an acute impact on human skin, leading to edema, erythema, or sunburn before pigmentation begins.²¹ In this

study, the erythema index and skin lightness index were observed by assessing the difference in the a^* value and the L^* value at the baseline to 12 hours post-UVB exposure. The higher a^* value indicates the increasingly erythematous state of the skin, while the higher L^* value indicates brighter skin.

Erythema response is influenced by the strength of the energy exposure and the presence or absence of protection. Erythema is usually seen 3 to 5 hours after UVB exposure and reaches maximum intensity 12 to 24 hours after exposure. Various biochemical courses at the cellular level occur as an initial response of the skin to UV radiation, which includes vasodilation of blood vessels, activation of endothelial cells, and increased levels of inflammatory mediators.^{3,22}

Excessive exposure to UVB radiation in the skin increases COX-2 protein receptors and the formation of reactive oxygen species (ROS), which triggers the expression of MMP1 that degrades collagen.²³ UVB exposure also results in massive neutrophil infiltration, in which the neutrophil will produce cytokines and chemokines that also activate MMP.²⁴ Erythema results from rapid degranulation of mast cells followed by TNF- α release and vasodilation of blood vessels.²² Therefore, the inflammatory process and erythema can be limited by reducing the formation of ROS, mast cells, and neutrophils or other inflammatory mediators.²⁵

The biochemical composition of MPE has been reported to contain flavonoids, which play an important role in counteracting free radicals, and xanthenes, which have various antibacterial, antifungal, and anti-inflammatory properties.²⁶ Of all the xanthenes that have been identified, α -mangostin has the highest level of xanthenes isolated from mangosteen pericarp.^{9,10}

High antioxidant activity on mangosteen pericarp xanthenes has been reported by various studies. Yoshikawa et al²⁷ found that methanol extract from mangosteen pericarp showed DPPH radical scavenging activity. Leong and Shui²⁸ compared the total antioxidant capacity using ABTS and DPPH tests, which describe a high antioxidant activity of mangosteen extract. In a similar study, Garcia et al²⁹ studied antioxidant capacity through the measurement of lipoperoxidation and hydroxyl radical scavenging activity, which reported that MPE had the highest antioxidant activity. Moongkarndi et al³⁰ also reported mangosteen extract significantly

TABLE 3. The Erythema Index (Δa^*) of each treatment site ($a^*_{24\text{hours}} - a^*_{\text{baseline}}$).

TREATMENT GROUPS	n	ERYTHEMA INDEX (Δa^*)			P-VALUE [†]
		MEDIAN	MIN	MAX	
Cream 5% + UVB	31	1.03 ^d	-1.09	5.25	<0.05
Cream 10% + UVB	31	-0.07 ^{ab}	-2.89	2.95	
Cream 20% + UVB	31	0.16 ^{ac}	-1.77	2.54	
Cream base + UVB	31	2.03 ^a	-0.69	6.22	
UVB	31	5.585 ^f	0.27	6.84	
No treatment	31	-0.15 ^{bc}	-1.27	0.795	

[†] $P < 0.05$ considered statistically significant (Kruskal-Wallis test)

^{abcd} similar superscript letters are not significantly different between groups (Mann-Whitney test)

TABLE 4. The Lightness Index (ΔL^*) of each treatment site ($L^*_{24\text{hours}} - L^*_{\text{baseline}}$).

TREATMENT GROUPS	n	LIGHTNESS INDEX (ΔL^*)			P-VALUE [†]
		MEDIAN	MIN	MAX	
Cream 5% + UVB	31	1.65 ^a	-5.4	5.29	<0.05
Cream 10% + UVB	31	-0.4 ^{bc}	-5.11	3.28	
Cream 20% + UVB	31	0.01 ^{bd}	-4.32	5.55	
Cream base + UVB	31	2.21 ^a	-1.85	6.83	
UVB	31	4.53 ^c	-0.015	6.755	
No treatment	31	-0.32 ^{cd}	-2.55	2.875	

[†] $P < 0.05$ considered statistically significant (Kruskal-Wallis test)

^{abcd} similar superscript letters are not significantly different between groups (Mann-Whitney test)

reduced intracellular ROS production through measurement of 2,2'-DCFH-DA. Jung et al³¹ also reported a high capacity of scavenging free radicals by γ -mangostin, gartanin, α -mangostin, garcinone B, garcimangone B, 1-isomangostin and garcinone D. Also, a study by Chomnawang et al³² measured antioxidant activity by inhibiting DPPH radical formation, which showed that mangosteen ethanol extract had significant antioxidant activity. Data from these various studies show the high antioxidant activity of MPE.⁷

In addition to being an antioxidant, various studies have also reported the role of α -mangostin in inhibiting inflammatory mediators.³³ Studies show that α -mangostin can lower the expression of inflammatory mediators induced by lipopolysaccharides such as TNF- α and IL-6.^{34,35} Inhibition of activation of MAPK, NF- κ B, AP-1 and diminishing of pro-inflammatory cytokine gene expression was also observed in other studies of α -mangostin treatment.^{36,37} Another study examined the inhibitory effect of α -mangostin on the secretion of pro-inflammatory mediators, where α -mangostin inhibits the secretion of IL-8 and TNF- α .³³ Other studies also evaluate the inhibition of α - and γ -mangostin against NO and PGE₂.^{38,39} This is

consistent with other studies that reported a dose-dependent γ -mangostin inhibiting PGE₂ release.⁴⁰ Similar studies report suppression of histamine release by α -, β - and γ -mangostin which is observed in rat basophilic leukemia cells.⁴¹ This effect is associated with decreased COX-2 mRNA and protein expression, and activation of NF- κ B. Other studies observed intraperitoneal and oral administration of α -mangostin, 1-isomangostin, or triacetate mangostin and reported an anti-inflammatory activity in several mouse inflammatory models.⁴² The *in-vivo* anti-inflammatory activity of γ -mangostin has been confirmed in edema models of the soles of rat feet.^{38,43} Administration of α and γ -mangostin orally also shows anti-inflammatory activity in ovalbumin-induced asthma (OVA) mice.⁴⁴ The topical administration of MPE gel has been reported to reduce periodontal inflammation.⁴⁵ Data from these studies show the anti-inflammatory roles of MPE through its bioactive content.

Antioxidant and anti-inflammatory effects are the main mechanism of the photoprotective activity of herbal extracts. Phenolic and flavonoid organic compounds are the main source of natural antioxidant compounds that can provide oxidative protection against free radicals

including ROS. In UV-damaged skin, where ROS is formed, ROS reacts negatively with DNA, protein, and fatty acids which can induce carcinogenic processes and inflammatory responses.⁴⁶

Several *in-vitro* studies have reported a fraction of the MPE compound having UV filter ability. This is based on a spectrophotometric assessment of the ability of these compounds to absorb light.⁴⁶ A study in Sri Lanka reported *in-vitro* research on the sunscreen activity of MPE with the highest concentration in the methanol fraction reaching an SPF value of 33.80.⁴⁷ This shows that the observed solar filtering effect is natural, intrinsic and has high potential to be developed into an inexpensive and efficacious and environmentally friendly sunscreen. Two other studies from Indonesia also reported the same thing. The first study reported the results of *in-vitro* testing of dichloromethane and butanol fractions of MPE that has a high UV-protection ability by reaching an SPF value >50.⁴⁸ The second study reported the ethanol fraction of MPE showed sunscreen activity that reached an SPF value >30 at a concentration of 100ppm.⁴⁹

CONCLUSION

In this study, the base cream group was found to protect against erythema and affect skin brightness 12 hours after UVB exposure. The base cream used contains titanium dioxide (TiO₂), which is a snow-white inorganic filter compound that is insoluble in water. TiO₂, with a size of more than 200nm, can protect the skin physically from all UV wavelengths.⁵⁰ However, the 10% and 20% MPE cream demonstrate a higher protective effect against UVB and was statistically significant. This study involved subjects with limited skin types; hence, additional research in other skin types would be beneficial in understanding the efficacy and tolerability.

REFERENCES

- Tobin DJ. Introduction to skin aging. *J Tissue Viability*. 2017;26(1):37–46.
- Buckman S, Gresham A, Hale P, et al. COX-2 expression is induced by UVB exposure in human skin: implications for the development of skin cancer. *Carcinogenesis*. 1998;19(5):723–729.
- Kochevar I, Taylor C, Krutmann J. Fundamentals of cutaneous photobiology and photoimmunology. *Fitzpatrick's dermatology in general medicine*. 2008;1:1031–1048.
- Im AR, Kim YM, Chin YW, et al. Protective effects of compounds from *Garcinia mangostana* L. (mangosteen) against UVB damage in HaCaT cells and hairless mice. *Int J Mol Med*. 2017;40(6):1941–1949.
- Gondokesumo ME, Pardjianto B, Sumitro SB, et al. Microstructural Characterization of the *Garcinia mangostana* Fruit at Different Maturity Level. *J Nat Med*. 2018;18(2):63–70.
- Chen G, Li Y, Wang W, et al. Bioactivity and pharmacological properties of α -mangostin from the mangosteen fruit: a review. *Expert Opin Ther Pat*. 2018;28(5):415–427.
- Pedraza-Chaverri J, Cárdenas-Rodríguez N, Orozco-Ibarra M, et al. Medicinal properties of mangosteen (*Garcinia mangostana*). *Food Chem Toxicol*. 2008;46(10):3227–3239.
- Obolskiy D, Pischel I, Siriwatanametanon N, et al. *Garcinia mangostana* L.: a phytochemical and pharmacological review. *Phytother Res*. 2009;23(8):1047–1065.
- Kurose H, Shibata MA, Iinuma M, et al. Alterations in cell cycle and induction of apoptotic cell death in breast cancer cells treated with α -mangostin extracted from mangosteen pericarp. *J Biomed Biotechnol*. 2012;2012:672428.
- Walker EB. HPLC analysis of selected xanthenes in mangosteen fruit. *J Sep Sci*. 2007;30(9):1229–1234.
- Marzaimi IN, Aizat WM. Current Review on Mangosteen Usages in Antiinflammation and Other Related Disorders. *Bioactive Food as Dietary Interventions for Arthritis and Related Inflammatory Diseases*: Elsevier; 2019. 273–289.
- American Academy of Dermatology Association statement on sunscreen access [press release]. ROSEMONT III: American Academy of Dermatology Association. Updated May 4, 2018.
- Seok JK, Kwak JY, Choi GW, et al. *Scutellaria radix* Extract as a Natural UV Protectant for Human Skin. *Phytother Res*. 2016;30(3):374–379.
- Runger T. Cutaneous Photobiology. In: Kang S, Amagai M, Bruckner A, Enk A, McMichael A, Orringer J, et al., editors. *Fitzpatrick's Dermatology*. 9th ed: McGraw-Hill Education; 2019. 272.
- Jaleel T, Pollack B, Elmets C. Phototherapy. In: Kang S, Amagai M, Bruckner A, et al., editors. *Fitzpatrick's Dermatology*. 9th ed: McGraw-Hill Education; 2019. 3640.
- Heckman CJ, Chandler R, Kloss JD, et al. Minimal Erythema Dose (MED) testing. *J Vis Exp*. 2013;(75):e50175. 2013.
- Jocher A, Kessler S, Hornstein S, et al. The UV erythema test as a model to investigate the anti-inflammatory potency of topical preparations—reevaluation and optimization of the method. *Skin Pharmacol Physiol*. 2005;18(5):234–240.
- Hughes-Formella BJ, Bohnsack K, Rippe F, et al. Anti-inflammatory effect of hamamelis lotion in a UVB erythema test. *Dermatology*. 1998;196(3):316–322.
- Del Bino S, Bernerd F. Variations in skin colour and the biological consequences of ultraviolet radiation exposure. *Br J Dermatol*. 2013;169 Suppl 3:33–40.
- Harrison GI, Young AR. Ultraviolet radiation-induced erythema in human skin. *Methods*. 2002;28(1):14–19.
- Young AR, Claveau J, Rossi AB. Ultraviolet radiation and the skin: Photobiology and sunscreen photoprotection. *J Am Acad Dermatol*. 2017;76(3S1):S100–S109.
- Clydesdale GJ, Dandie GW, Muller HK. Ultraviolet light induced injury: immunological and inflammatory effects. *Immunol Cell Biol*. 2001;79(6):547–568.
- Marwaha V, Chen Y-H, Helms E, et al. T-oligo treatment decreases constitutive and UVB-induced COX-2 levels through p53-and NF κ B-dependent repression of the COX-2 promoter. *J Biol Chem*. 2005;280(37):32379–32388.
- Han YP, Tuan TL, Wu H, et al. TNF-alpha stimulates activation of pro-MMP2 in human skin through NF-(kappa)B mediated induction of MT1-MMP. *J Cell Sci*. 2001;114(Pt 1):131–139.
- Che DN, Xie GH, Cho BO, et al. Protective effects of grape stem extract against UVB-induced damage in C57BL mice skin. *J Photochem Photobiol B*. 2017;173:551–559.
- Pothitirat W, Chomnawang MT, Supabphol R, et al. Comparison of bioactive compounds content, free radical scavenging and anti-acne inducing bacteria activities of extracts from the mangosteen fruit rind at two stages of maturity. *Fitoterapia*. 2009;80(7):442–447.
- Yoshikawa M, Harada E, Miki A, et al. Antioxidant constituents from the fruit hulls of mangosteen (*Garcinia mangostana* L.) originating in Vietnam. *Journal of the Pharmaceutical Society of Japan*. 1994;114(2):129–133.
- Leong L, Shui G. An investigation of antioxidant capacity of fruits in Singapore markets. *Food Chem*. 2002;76(1):69–75.
- Garcia VV, Magpantay TO, Escobin LD. Antioxidant potential of selected Philippine vegetables and fruits. *Philippine Agricultural Scientist*. 2005;88(1):78.
- Moongkarndi P, Kosem N, Luanratana O, et al. Antiproliferative activity of Thai medicinal plant extracts on human breast adenocarcinoma cell line. *Fitoterapia*. 2004;75(3-4):375–377.
- Jung H-A, Su B-N, Keller WJ, Mehta RG, Kinghorn AD. Antioxidant xanthenes from the pericarp of *Garcinia mangostana* (Mangosteen). *J Agric Food Chem*. 2006;54(6):2077–2082.

32. Chomnawang MT, Surassmo S, Nukoolkarn VS, Gritsanapan W. Effect of *Garcinia mangostana* on inflammation caused by *Propionibacterium acnes*. *Fitoterapia*. 2007;78(6):401–408.
33. Gutierrez-Orozco F, Chitchumroonchokchai C, Lesinski GB, et al. α -Mangostin: anti-inflammatory activity and metabolism by human cells. *J Agric Food Chem*. 2013;61(16):3891–3900.
34. Bumrungpert A, Kalpravidh RW, Chuang C-C, et al. Xanthones from mangosteen inhibit inflammation in human macrophages and in human adipocytes exposed to macrophage-conditioned media. *J Nutr*. 2010;140(4):842–847.
35. Liu S-H, Lee L-T, Hu N-Y, et al. Effects of alpha-mangostin on the expression of anti-inflammatory genes in U937 cells. *Chin Med*. 2012;7(1):19.
36. Bumrungpert A, Kalpravidh RW, Chitchumroonchokchai C, Chuang C-C, West T, et al. Xanthones from mangosteen prevent lipopolysaccharide-mediated inflammation and insulin resistance in primary cultures of human adipocytes. *J Nutr*. 2009;139(6):1185–1191.
37. Bumrungpert A, Kalpravidh RW, Suksamrarn S, et al. Bioaccessibility, biotransformation, and transport of α -mangostin from *Garcinia mangostana* (Mangosteen) using simulated digestion and Caco-2 human intestinal cells. *Molecular Nutrition & Food Research*. 2009;53(S1):S54–S61.
38. Chen L-G, Yang L-L, Wang C-C. Anti-inflammatory activity of mangostins from *Garcinia mangostana*. *Food Chem Toxicol*. 2008;46(2):688–693.
39. Tewtrakul S, Wattanapiromsakul C, Mahabusarakam W. Effects of compounds from *Garcinia mangostana* on inflammatory mediators in RAW264.7 macrophage cells. *J Ethnopharmacol*. 2009;121(3):379–382.
40. Nakatani K, Nakahata N, Arakawa T, et al. Inhibition of cyclooxygenase and prostaglandin E2 synthesis by γ -mangostin, a xanthone derivative in mangosteen, in C6 rat glioma cells. *Biochem Pharmacol*. 2002;63(1):73–79.
41. Itoh T, Ohguchi K, Iinuma M, et al. Inhibitory effect of xanthones isolated from the pericarp of *Garcinia mangostana* L. on rat basophilic leukemia RBL-2H3 cell degranulation. *Bioorg Med Chem*. 2008;16(8):4500–4508.
42. Shankaranarayan D, Gopalakrishnan Ct, Kameswaran L. Pharmacological profile of mangostin and its derivatives. *Archives internationales de pharmacodynamie et de therapie*. 1979;239(2):257–269.
43. Nakatani K, Yamakuni T, Kondo N, et al. γ -Mangostin inhibits inhibitor- κ B kinase activity and decreases lipopolysaccharide-induced cyclooxygenase-2 gene expression in C6 rat glioma cells. *Mol Pharmacol*. 2004;66(3):667–674.
44. Jang H-Y, Kwon O-K, Oh S-R, et al. Mangosteen xanthones mitigate ovalbumin-induced airway inflammation in a mouse model of asthma. *Food Chem Toxicol*. 2012;50(11):4042–4050.
45. Rassameemasmaung S, Sirikulsathean A, Amornchat C, et al. Topical application of *Garcinia mangostana* L. pericarp gel as an adjunct to periodontal treatment. *Complement Ther Med*. 2008;16(5):262–267.
46. Buso P, Radice M, Baldisserotto A, et al. Guidelines for the Development of Herbal-Based Sunscreen. *Herbal Medicine: IntechOpen*; 2017.
47. Ratnasooriya W, Pathirana R, Gamage R, et al. Sunscreen Activity of Pericarp of Fruit of Sri Lankan *Garcinia mangostana* L. (Mangosteen) in vitro. *IMP*. 2017;3.
48. Susanti M, Putra DP. In-vitro UV protection activity of *Garcinia mangostana*. *Pharmaceutical Journal of Indonesia*. 2012;13(2):61–64.
49. Iwo MI, Soemardji AA, Hanafi M. Sunscreen activity of α -mangostin from the pericarps of *Garcinia mangostana* Linn. *J Appl Pharm Sci*. 2013;3(6):70–73.
50. Chung J. Photoprotection. In: Kang S, Amagai M, Bruckner A, et al., editors. *Fitzpatrick's Dermatology*. 9th ed: McGraw-Hill Education; 2019. 3623. **JCAD**